

AMERICAN WATER RESOURCES ASSOCIATION

# GLYPHOSATE AND ITS DEGRADATION PRODUCT AMPA OCCUR FREQUENTLY AND WIDELY IN U.S. SOILS, SURFACE WATER, GROUNDWATER, AND PRECIPITATION<sup>1</sup>

W.A. Battaglin, M.T. Meyer, K.M. Kuivila, and J.E. Dietze<sup>2</sup>

ABSTRACT: Glyphosate use in the United States increased from less than 5,000 to more than 80,000 metric tons/ yr between 1987 and 2007. Glyphosate is popular due to its ease of use on soybean, cotton, and corn crops that are genetically modified to tolerate it, utility in no-till farming practices, utility in urban areas, and the perception that it has low toxicity and little mobility in the environment. This compilation is the largest and most comprehensive assessment of the environmental occurrence of glyphosate and aminomethylphosphonic acid (AMPA) in the United States conducted to date, summarizing the results of 3,732 water and sediment and 1,018 quality assurance samples collected between 2001 and 2010 from 38 states. Results indicate that glyphosate and AMPA are usually detected together, mobile, and occur widely in the environment. Glyphosate was detected without AMPA in only 2.3% of samples, whereas AMPA was detected without glyphosate in 17.9% of samples. Glyphosate and AMPA were detected frequently in soils and sediment, ditches and drains, precipitation, rivers, and streams; and less frequently in lakes, ponds, and wetlands; soil water; and groundwater. Concentrations of glyphosate were below the levels of concern for humans or wildlife; however, pesticides are often detected in mixtures. Ecosystem effects of chronic low-level exposures to pesticide mixtures are uncertain. The environmental health risk of lowlevel detections of glyphosate, AMPA, and associated adjuvants and mixtures remain to be determined.

(KEY TERMS: glyphosate; AMPA; water quality; surface water; groundwater; precipitation.)

Battaglin, W.A., M.T. Meyer, K.M. Kuivila, and J.E. Dietze, 2014. Glyphosate and Its Degradation Product AMPA Occur Frequently and Widely in U.S. Soils, Surface Water, Groundwater, and Precipitation. *Journal of the American Water Resources Association* (JAWRA) 50(2): 275-290. DOI: 10.1111/jawr.12159

### INTRODUCTION

### Problem

Commercial glyphosate [N-(phosphonomethyl) glycine] formulations have been used worldwide for decades, but glyphosate is seldom included in environmental monitoring programs (Gilliom *et al.*, 2006; Loos *et al.*, 2010; U.S. Department of Agriculture,

2011), due in part to difficulties in quantifying this polar and water-soluble compound at environmentally relevant concentrations (Skark *et al.*, 1998; Sanchis *et al.*, 2011). In the early 2000s, scientists at the U.S. Geological Survey (USGS) began developing analytical methods (Lee *et al.*, 2002) and conducting reconnaissance studies (Scribner *et al.*, 2003; Battaglin *et al.*, 2005) for the occurrence of glyphosate and aminomethylphosphonic acid (AMPA) in anticipation of growing gaps in scientific understanding due to (1)

<sup>1</sup>Paper No. JAWRA-13-0028-P of the *Journal of the American Water Resources Association* (JAWRA). Received February 27, 2013; accepted June 18, 2013. © 2014 American Water Resources Association. This article is a U.S. Government work and is in the public domain in the USA. **Discussions are open until six months from print publication**.

<sup>&</sup>lt;sup>2</sup>Respectively, Research Hydrologist (Battaglin), U.S. Geological Survey Colorado Water Science Center, Box 25046, MS 415, DFC, Lakewood, Colorado 80225; Supervisory Research Chemist (Meyer), U.S. Geological Survey Kansas Water Science Center, Lawrence, Kansas 66049; Research Hydrologist (Kuivila), U.S. Geological Survey Oregon Water Science Center, Portland, Oregon 97201; and Physical Science Technician (Dietze), U.S. Geological Survey Kansas Water Science Center, Lawrence, Kansas 66049 (E-Mail/Battaglin: wbattagl@usgs.gov).

the widespread agricultural and nonagricultural use of glyphosate, (2) the rapid increase in glyphosate use starting in 1997 corresponding to the introduction of genetically modified glyphosate-resistant crops like soybeans and corn, and (3) the absence of information on the environmental occurrence of glyphosate and AMPA. These USGS scientists continued to develop new analytical methods and began a series of studies to determine the fate of glyphosate and AMPA in the environment. Researchers from Canada also have noted the need for methods to monitor glyphosate due to its increasing use (Byer *et al.*, 2008).

#### Study Objective

The objective of this investigation was to broadly summarize glyphosate and AMPA occurrence and concentration in water and sediment samples collected in 2001 through 2010 from diverse hydrologic settings and a wide geographic range of locations in the United States (U.S.). The data also are used to identify, in which hydrologic settings glyphosate and AMPA are more or less likely to occur and to a limited degree the temporal patterns of their occurrence or concentrations over the study period. The data used in this analysis were collected by a series of studies (Scribner et al., 2003, 2007; Kolpin et al., 2004, 2006; Battaglin et al., 2005, 2009; Baker et al., 2006; McCarthy et al., 2011; Coupe et al., 2012), most, but not all of which were designed to determine the fate of glyphosate and AMPA or other pesticides in the environment.

#### Glyphosate Use

Herbicides containing glyphosate are used in more than 130 countries on more than 100 crops (Monsanto, 2009). Glyphosate was first registered for use in the U.S. in 1974 in Roundup<sup>®</sup> and is the most heavily used pesticide for agriculture, and the second most heavily used pesticide for home and garden and commercial/industrial sectors in the U.S. Glyphosate use in U.S. agriculture has increased dramatically from  $\sim$ 3,180 metric tons of active ingredient in 1987 to ~82,800 metric tons in 2007 (Figure 1) (Kiely et al., 2004; Grube et al., 2011). Glyphosate accounted for about 40% of all herbicide use (by weight of active ingredient) in the U.S. in 2007. Similar increases in glyphosate use also have occurred in Canada (Struger et al., 2008). The majority of this increase is the result of glyphosate use on soybean, cotton, canola, and corn crops that have been genetically modified to tolerate this glyphosate (e.g., Roundup<sup>®</sup> Ready crops) (Cerdeira and Duke, 2006; Young, 2006). About 80% of all genetically modified crops planted worldwide are designed to

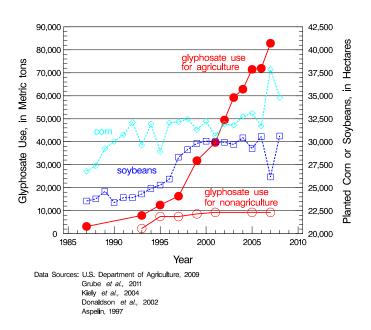


FIGURE 1. Use in U.S. of Glyphosate and Planted Hectares of Corn and Soybeans, 1987-2008.

tolerate glyphosate (Dill *et al.*, 2008), hence these crops comprise the "overwhelming majority" of herbicideresistant crops (Benbrook, 2012). Glyphosate is typically (but not always) applied "post-emergence" or after crops and weeds have emerged from the soil, and may be applied more than once during a growing season. Glyphosate use also has increased due to increased use of "no-till" farming practices on crops that are not genetically modified to tolerate glyphosate (Horowitz *et al.*, 2010). Glyphosate loading rates (total use in a county divided by county land area) are largest in the corn and soybean producing region of the Midwest, along the Mississippi River alluvial floodplain, and in parts of California and Florida (Figure 2).

Glyphosate is popular with farmers for a number of reasons. Some studies indicate that the planting of glyphosate-tolerant crops in U.S. agriculture has saved farmers money and reduced the total pounds of herbicides applied (Gianessi and Sankula, 2003; Gianessi, 2008). Another reason for its popularity is the perception that glyphosate is an "environmentally benign" herbicide (Giesy *et al.*, 2000; Duke and Powles, 2008) that has low toxicity and little mobility or persistence in the environment. However, other studies indicate that glyphosate-resistant weeds can become a problem in areas where glyphosate-based crop production systems are used (Owens, 2008; Powles, 2008).

Glyphosate is also commonly used by homeowners and for other nonagricultural purposes. The nonagricultural use of glyphosate has increased from 2,270 metric tons in 1993 to 9,300 metric tons in 2007 (Figure 1) (Aspelin, 1997; Kiely *et al.*, 2004; Grube *et al.*, 2011). Urban glyphosate use can result in contamination of areas downstream from wastewa-

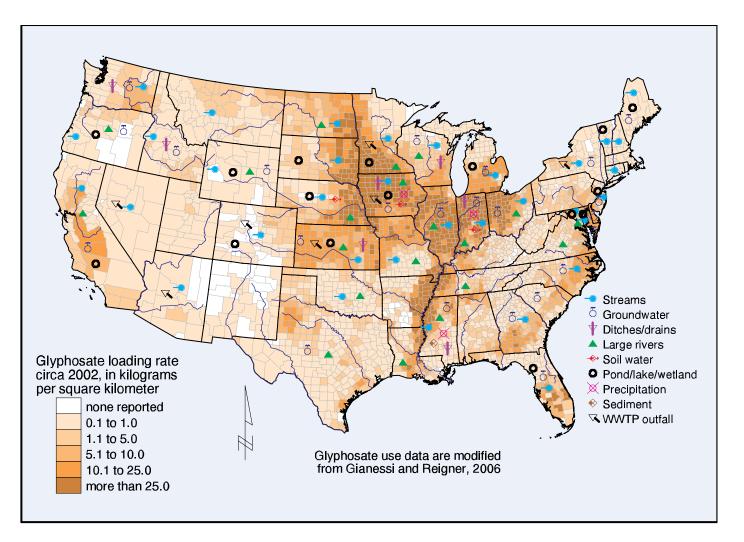


FIGURE 2. Glyphosate Loading Rate, Circa 2002 (the most recent year for which county-scale estimates of glyphosate sales are publicly available), and State in Which Various Hydrologic Settings Were Sampled and Analyzed for Glyphosate and AMPA (actual site locations are not shown). WWTP, wastewater treatment plant.

ter treatment plants (WWTPs) or storm sewers (Kolpin *et al.*, 2006; Botta *et al.*, 2009; Hanke *et al.*, 2010). Although pesticide application amounts in urban areas are generally less than in agricultural regions, those applications are frequently on or near impervious surfaces and can result in substantial pesticide inputs to urban drainage systems (Blanchoud *et al.*, 2007; Wittmer *et al.*, 2011).

# Toxicity

Glyphosate is a nonselective contact herbicide that kills plants by inhibiting the synthesis of aromatic amino acids needed for protein formation (Franz *et al.*, 1997). Glyphosate is no more than slightly toxic to birds, fish, and aquatic invertebrates and exhibits low oral and dermal acute toxicity to humans (U.S. Environmental Protection Agency, 1993). Glyphosate showed little effect on soil microbial communities (Haney *et al.*, 2000; Busse *et al.*, 2010) and limited effect on riverine microbial communities at exposures of about 10  $\mu$ g/l (Pesce *et al.*, 2009).

Recent studies, however, have documented the potential for sublethal and other toxic effects of glyphosate and its adjuvant formulations. A recent study suggests that glyphosate exposure can delay periphytic colonizations, reduce diatom abundance, and enhance the development of cyanobacteria in shallow lakes (Vera et al., 2010). Some research suggests that glyphosate, at environmentally realistic concentrations, can act synergistically with parasites to reduce fish survival (Kelly *et al.*, 2010). Glyphosate also seems to cause malformations by interfering with retinoic acid signaling in Xenopus laevis (Paganelli et al., 2010). At least one study has suggested that glyphosate-based herbicides are "info-disruptors" that can interfere with chemical communications between male and female spiders (Griesinger et al., 2011). Other research suggests that glyphosate can

negatively impact microbial activity in the root zone of glyphosate-resistant soybeans (Zobiole *et al.*, 2010) resulting in reduced plant growth and reduced resistance to pathogen colonization. Interestingly, glyphosate exposure appeared to reduce *Batrachochytrium dendrobatidis* (Bd) caused mortality in exposed wood frogs, presumably having a greater adverse effect on the pathogen than the host (Gahl *et al.*, 2011). Glyphosate is on the U.S. Environmental Protection Agency list of pesticide active ingredients that will be tested for potential hormonal effects under its Endocrine Disruptor Screening Program (U.S. Environmental Protection Agency, 2009).

AMPA acid is generally considered to be less toxic, or of no greater toxicological concern, than glyphosate (FAO, 1997; Giesy et al., 2000), however, few studies have done direct comparisons of the toxicity of glyphosate and AMPA on non-target species. From the review by Giesy et al. (2000), AMPA was equally toxic as glyphosate to green algae (Scenedesmus subspicatus), equally toxic to birds (Colinus virginianus), equally toxic to terrestrial mammals (rat), slightly more toxic to aquatic invertebrates (Daphnia magna), and substantially less toxic to fish (Oncorhynchus mykiss). AMPA was relatively toxic to fungus (Glomus *intraradices*) but less so than glyphosate (Wan *et al.*, 1998). AMPA was found to have a clastogenic effect in human lymphocytes and otherwise demonstrated genotoxicity using the Comet assay (Mañas et al., 2009). The tolerances established by the U.S. Environmental Protection Agency under 40 CFR Part 180 for commodities such as grains or livestock are "expressed in terms of glyphosate, including its metabolites and degradates" (U.S. Environmental Protection Agency, 1993, 2011).

Some studies indicate that commercial glyphosate formulations can be more toxic than pure glyphosate due to the toxicity and (or) action of the surfactants and other adjuvants used (Giesy et al., 2000; Edginton et al., 2004; Bringolf et al., 2007; Mesnage et al., 2012; Moore *et al.*, 2012). Surfactants such as polyethoxylated tallowamine (POEA) are added to some commercial glyphosate formulations to enhance its efficacy. The Roundup<sup>®</sup> formulation was more toxic than glyphosate or AMPA for all taxa tested (Giesy et al., 2000). Some formulations of POEA were toxic to Daphnia magna, inhibiting growth and causing mortality at concentrations less than 100 µg/l (Brausch et al., 2007). Effects on the development and survival of amphibians have been observed at various levels of glyphosate and POEA exposure (Lajmanovich et al., 2003; Edginton et al., 2004; Howe et al., 2004; Cauble and Wagner, 2005; Relyea, 2005a, b, 2012; Dinehart et al., 2009; Mann et al., 2009; Jones et al., 2010; King and Wagner, 2010; Lenkowski et al., 2010; Williams and Semlitsch, 2010; Moore et al., 2012).

Glyphosate and several glyphosate formulations have a cytotoxic effect on human cells, and endocrine disruption, specifically inhibition of estrogen synthesis, has been demonstrated (Richard *et al.*, 2005; Benachour *et al.*, 2007; Benachour and Seralini, 2009; Mesnage *et al.*, 2012). Glyphosate formulations also may cause birth defects or adverse reproductive effects in vertebrates or contribute to a variety of human diseases (Daruich *et al.*, 2001; Dallegrave *et al.*, 2003, 2007; Paganelli *et al.*, 2010; Samsel and Seneff, 2013).

# Environmental Fate

Glyphosate is a polar, amphoteric compound that binds strongly to soils, but also is very water soluble (more than 10,000 mg/l at 25°C). Glyphosate has a soil half-life that ranges from 2 to 215 days, and an aquatic half-life that ranges from 2 to 91 days (Giesv *et al.*.. 2000; Grunewald et al., 2001; National Pesticide Information Center, 2008; Vera et al., 2010). Glyphosate degrades in the environment, primarily by microbial processes, to AMPA. AMPA also is very water soluble, and it degrades more slowly than glyphosate (Grunewald et al., 2001). AMPA has a soil half-life that ranges from 60 to 240 days and an aquatic half-life that is comparable to that of glyphosate (Giesy *et al.*, 2000; Bergstrom et al., 2011). AMPA ultimately degrades to inorganic phosphate, ammonium, and  $CO_2$  (Borggaard and Gimsing, 2008), a process that can result in substantial increases in total phosphorous in aquatic systems (Vera et al., 2010). Glyphosate also can be degraded by bacteria to sarcosine but this process has not been well documented in soils (Borggaard and Gimsing, 2008). AMPA also can be formed by the degradation of phosphonic acids found in some household and industrial detergents and cleaning products (Skark et al., 1998; Nowack, 2003) making outfall from WWTPs and septic tanks a potential source of AMPA in some areas (Kolpin et al., 2006; Botta et al., 2009). However, phosphonic acids are strongly adsorbed to sediments and suspended particles, and recalcitrant to biological or non-biological degradation (HERA, 2004).

## METHODS

### Study Sites and Sample Collection

A total of 3,732 environmental samples collected from 38 states and the District of Columbia were included in this analysis. The hydrologic settings sampled include groundwater, streams (having drain-

| Hydrologic<br>Setting         | Number of<br>Samples | Percentage and<br>(number) with<br>Glyphosate<br>Detections | Median<br>Glyphosate<br>in µg/l or µg/kg | Maximum<br>Glyphosate<br>in µg/l or µg/kg | Percentage and<br>(number) with<br>AMPA Detections | Median<br>AMPA<br>in µg/l or<br>µg/kg | Maximum<br>AMPA in µg/l<br>or µg/kg |
|-------------------------------|----------------------|---|--|---|--|---------------------------------------|-------------------------------------|
| All sites                     | 3,732                | 39.4 (1,470)  | < 0.02                                   | 476                                       | 55.0 (2,052)                                       | 0.04                                  | 397                                 |
| Streams                       | 1,508                | 52.5 (791)  | 0.03                                     | 73  | 71.6 (1,079)                                       | 0.20                                  | 28                                  |
| Groundwater                   | 1,171                | 5.8 (68)  | $<\!\!0.02$                              | 2.03                                      | 14.3 (168)   | < 0.02                                | 4.88                                |
| Ditches and<br>drains         | 374                  | 70.9 (265)  | 0.20                                     | 427                                       | 80.7 (302)   | 0.43                                  | 397                                 |
| Large rivers                  | 318                  | 53.1 (169)  | 0.03                                     | 3.08                                      | 89.3 (284)   | 0.22                                  | 4.43                                |
| Soil water                    | 116                  | 34.5 (40)   | $<\!\!0.02$                              | 1.00                                      | 65.5 (76)  | 0.06                                  | 1.91                                |
| Lakes, ponds,<br>and wetlands | 104                  | 33.7 (35)   | < 0.02                                   | 301                                       | 29.8 (31)  | < 0.02                                | 41                                  |
| Precipitation                 | 85                   | 70.6 (60)   | 0.11                                     | 2.50                                      | 71.8 (61)  | 0.04                                  | 0.48                                |
| Soil and sediment             | 45                   | 91.1 (41)   | 9.6                                      | 476                                       | 93.3 (42)  | 18.0                                  | 341                                 |
| WWTP outfall                  | 11                   | 9.09 (1)  | $<\!\!0.02$                              | 0.30                                      | 81.8 (9)   | 0.45                                  | 2.54                                |

| TABLE 1. Number of Samples, Percentage Detections, and Median and Maximum Glyphosat | e |
|---|---|
| and AMPA Concentrations by Hydrologic Setting.                                      |   |

age areas less than 10,000 km<sup>2</sup>), rivers (having drainage areas greater than 10,000 km<sup>2</sup>), ditches and drains (both tile and surface), lakes, ponds, and wetlands, precipitation, WWTP outfalls, soil water, and sediment (Figure 2, Table 1). The most samples were collected from streams (1,508) followed by groundwater (1,171); ditches and drains (374); rivers (318); soil water (116); lakes, ponds, and wetlands (104); precipitation (85); sediment (45); and WWTP outfalls (11).

## Analytical Methods

In 2000-2002, the USGS developed an analytical method (Lee et al., 2002) that used online solid-phase extraction (SPE), and liquid chromatography/mass spectrometry (LC/MS) for determination of concentrations of glyphosate and AMPA in water samples with a reporting level of  $0.1 \mu g/l$  for both compounds. The method was modified, beginning in April 2004, to use isotope dilution and online SPE and liquid chromatography/tandem mass spectrometry (LC/MS/MS),which improved sensitivity and lowered the reporting level to  $0.02 \mu g/l$ , for both compounds (Meyer *et al.*, 2009). The lower analytical reporting level made it possible for environmental researchers to gain a better understanding of the fate and transport of glyphosate and AMPA. In a few samples (seven for glyphosate and five for AMPA) concentrations less than the 0.02 µg/l reporting level were measured and reported.

## Statistical Methods

When glyphosate or AMPA concentrations were less than the reporting level, those concentrations were set to zero for the purposes of calculating detection frequencies, the total glyphosate concentration, or other statistics; and to the reporting level for plotting. The total glyphosate concentration was calculated as the sum of glyphosate and AMPA The relative percent difference concentrations. between two concentration values (e.g., laboratory duplicates) was calculated as the absolute value of the difference between the two concentrations divided by the maximum of the two concentrations, that quantity multiplied by 100. Estimates of the instantaneous total glyphosate daily flux were calculated for samples at selected sites. Instantaneous daily fluxes in grams per day (or in some cases kilograms per day) for the date of sample collection were estimated as the product of the total glyphosate concentration (micrograms per liter), daily mean discharge (cubic feet per second), and 2.4463 (a units conversion). Instantaneous daily total glyphosate fluxes were estimated as zero on days when both glyphosate and AMPA were not detected in a sample. The Wilcoxon signed-rank test was used to determine if differences between groups of data are statistically significant (Helsel and Hirsch, 2002). The percentage AMPA (%AMPA) was calculated as shown below (Equation 1) where [AMPA] and [glyphosate] are their respective concentrations in water. %AMPA was set to zero (0.01  $\mu$ g/l for plotting purposes) when glyphosate was detected and AMPA was not, and not calculated when both glyphosate and AMPA were not detected. This ratio gives some insight into sources, fate, and transport of glyphosate and AMPA in the environment.

$$\text{MAMPA} = ([AMPA]/([glyphosate] + [AMPA])) \times 100$$

## **Quality Assurance Samples**

A total of 1,018 quality assurance (QA) samples were collected and analyzed in conjunction with the

(1)

3,732 environmental (ENV) samples described here. QA samples consisted of 514 laboratory duplicates (or ~14% of ENV samples although some were duplicates of other QA samples), 288 field replicates (~7.7% of ENV samples), 188 field blanks (~5.0% of ENV samples), and 28 field spikes. Glyphosate was not detected in any of the 188 field blanks. AMPA was detected in 2 of 188 field blanks, both surface water sites, at concentrations of 0.02 and 0.04 µg/l (both of these samples had a reporting level of 0.02 µg/l).

In 514 laboratory duplicate sample pairs, the presence or absence of glyphosate and AMPA was confirmed in 96% of the sample pairs. Glyphosate was detected in both samples in 198 sample pairs and in one of the two samples in 18 sample pairs. The relative percent differences in these 216 sample pairs ranged from 0 to 100, and median and mean percent differences were 10.0 and 20.1, respectively. The absolute difference in measured concentrations between environmental samples and laboratory duplicates ranged from 0 to 58 µg/l, and median and mean differences were 0.03 and 0.38 µg/l, respectively. AMPA was detected in both samples in 273 sample pairs and in one of the two samples in 19 sample pairs. The relative percent differences in these 292 sample pairs ranged from 0 to 100, and median and mean percent differences were 9.9 and 19.2, respectively. The absolute difference in measured detections ranged from 0 to 55 µg/l, and median and mean absolute difference were 0.03 and  $0.29 \ \mu g/l$ , respectively.

In 288 field replicate sample pairs, the presence or absence of glyphosate was confirmed in 98% of sample pairs, whereas the presence or absence of AMPA was confirmed in 97% of sample pairs. Glyphosate was detected in both samples in 70 sample pairs and in one of the two samples in 6 sample pairs. The relative percent differences in these 76 sample pairs ranged from 0 to 100, and median and mean percent differences were 17.0 and 25.2, respectively. The absolute difference in measured detections between environmental samples and laboratory duplicates ranged from 0 to 27 µg/l, and median and mean absolute differences were 0.04 and 0.79  $\mu$ g/l, respectively. AMPA was detected in both samples in 113 sample pairs and in one of the two samples in 9 sample pairs. The relative percent differences in these 122 sample pairs ranged from 0 to 100, and median and mean percent differences were 14.6 and 23.1, respectively. The absolute difference in measured detections ranged from 0 to 26 µg/l, and median and mean absolute differences were 0.03 and 0.37  $\mu$ g/l, respectively. For both laboratory duplicates and field replicates. differences larger than 1  $\mu$ g/l were rare and were typically observed in samples with high (greater than 5  $\mu$ g/l) concentrations of glyphosate or AMPA. Results from 28 field spike samples were not analyzed for this report.

## RESULTS

A total of 3,732 water or sediment samples were collected from 1,341 sites in 38 states and the District of Columbia. Glyphosate was detected at least once in samples from 289 sites, whereas AMPA was detected at least once at 384 sites. Glyphosate was detected in 1,470 of 3,732 or 39.4% of all environmental samples, and AMPA was detected in 2,052 of 3,732 or 55.0% of all environmental samples (Table 1). The median and maximum glyphosate concentrations in all samples were <0.02 and  $476 \,\mu g/l$ , respectively. The median and maximum AMPA concentrations in all samples were 0.04 and 397 µg/l, respectively. Glyphosate was detected in more than 50% of samples of sediment, ditches and drains, precipitation, large rivers, and streams and in less than 40% of samples of lakes, ponds, and wetlands; soil water; WWTP outfalls; and groundwater (Table 1, Figure 3). AMPA was detected in more than 50% of samples of soil and sediment, large rivers, WWTP outfalls, ditches and drains, precipitation, streams, and soil water; and in less than 30% of samples of lakes, ponds, and wetlands; and groundwater (Table 1, Figure 3). It was uncommon for glyphosate to be detected without AMPA, happening in only 2.3% of all samples. AMPA was detected without glyphosate in 17.9% of all samples. Both

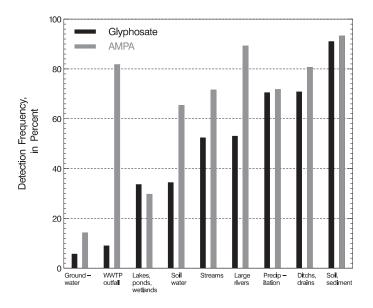


FIGURE 3. Detection Frequencies for Glyphosate and AMPA by Hydrologic Setting.

glyphosate and AMPA were detected in 37.1% of all samples, and neither glyphosate nor AMPA was detected in 42.7% of all samples.

## Soil and Sediment, and Soil Water

A total of 45 soil and sediment samples were collected from seven sites in Indiana and Mississippi (Figure 2). Glyphosate and AMPA were detected at least once in samples from all seven sites. Both glyphosate and AMPA were detected in more than 90% of sediment samples with concentrations frequently exceeding 10  $\mu$ g/kg (Figures 3 and 4). The median and maximum glyphosate concentrations in these samples were 9.6 and 476  $\mu$ g/kg, respectively, whereas the median and maximum AMPA concentrations were 18 and 341  $\mu$ g/kg, respectively. The median %AMPA ratio (in 42 samples) was 65% with an interquartile range of 55-78% (Figure 5).

A total of 116 soil water samples were collected from 13 sites in Indiana, Iowa, and Nebraska. Glyphosate was detected at least once in samples from nine sites, whereas AMPA was detected at least once at 12 sites. Glyphosate was detected in 34.5% and AMPA in 66.5% of soil water samples (Figures 3 and 4). The median and maximum glyphosate concentrations in these samples were <0.02 and 1.0  $\mu$ g/l, respectively, whereas the median and maximum AMPA concentrations were 0.06 and 1.91  $\mu$ g/l, respectively (Table 1). The median %AMPA ratio (in 79 samples) was 89% with an interquartile range of 76-100% (Figure 5).

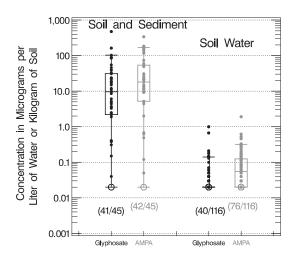


FIGURE 4. Boxplot-Dot Plots Showing Concentrations of Glyphosate and AMPA for Soil and Sediment and Soil Water Samples (numbers in parentheses are number of detections/number of samples, open circle is reporting level).

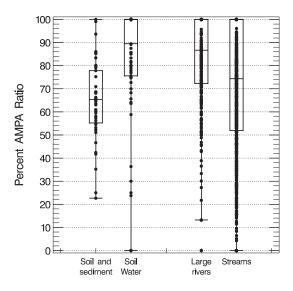


FIGURE 5. Boxplot-Dot Plots Showing the Percent AMPA Ratio for Soil and Sediment, Soil Water, Large River (drainage basin area 10,000 km<sup>2</sup> or greater), and Stream Samples.

## Large Rivers and Streams

A total of 318 large river (drainage basin area  $10,000 \text{ km}^2$  or greater at the sampling site) samples were collected from 47 sites in California, Iowa, Illinois, Indiana, Kansas, Louisiana, Maryland, Minnesota, Missouri, Mississippi, North Dakota, Nebraska, Ohio, Oklahoma, Oregon, Tennessee, Texas, Virginia, Wisconsin, and Wyoming. Glyphosate was detected at least once in samples from 32 sites, whereas AMPA was detected at least once at 42 sites. Glyphosate was detected in 53.1% and AMPA in 89.3% of large river samples (Figures 3 and 6). The median and maximum glyphosate concentrations in these samples were 0.03 and 3.08  $\mu$ g/l respectively, whereas the median and maximum AMPA concentrations were 0.22 and 4.43 µg/l, respectively (Table 1). The median %AMPA ratio (in 285 samples) was 87% with an interquartile range of 72-100% (Figure 5).

A total of 1,508 stream (drainage basin area less than 10,000 km<sup>2</sup> at the sampling site) samples were collected from 358 sites in Arizona, California, Colorado, Connecticut, District of Columbia, Florida, Georgia, Iowa, Idaho, Illinois, Indiana, Kansas, Maryland, Maine, Michigan, Minnesota, Missouri, Mississippi, Montana, North Dakota, Nebraska, New Hampshire, New Jersey, New York, Nevada, Ohio, Oklahoma, Oregon, South Dakota, Virginia, Vermont, Washington, Wisconsin, and Wyoming. Glyphosate was detected at least once in samples from 155 sites, whereas AMPA was detected at least once at 210 sites. Glyphosate was detected in 52.5% and AMPA in 71.6% of stream samples (Figures 3 and 6). The median and maximum glyphosate concentrations in

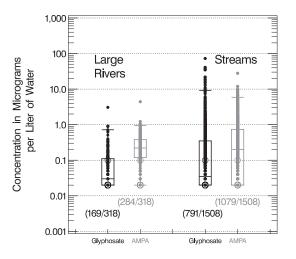


FIGURE 6. Boxplot-Dot Plots Showing Concentrations of Glyphosate and AMPA for Large River (drainage basin area  $10,000 \text{ km}^2$ or greater) and Stream Water Samples (numbers in parentheses are number of detections/number of samples, open circle is reporting level).

these samples were 0.03 and 73  $\mu$ g/l, respectively, whereas the median and maximum AMPA concentrations were 0.20 and 28  $\mu$ g/l, respectively (Table 1). The median %AMPA ratio (in 1,116 samples) was 74% with an interquartile range of 52-100% (Figure 5).

### Ditches and Drains; and Lakes, Ponds, and Wetlands

A total of 374 ditch, tile, or surface drain samples were collected from 32 sites in Iowa, Idaho, Indiana, Kansas, Mississippi, Washington, and Wisconsin. Glyphosate was detected at least once in samples from 23 sites, whereas AMPA was detected at least once at 24 sites. Glyphosate was detected in 70.9%, and AMPA in 80.7% of ditch or drain samples (Figures 3 and 7). The median and maximum glyphosate concentrations in these samples were 0.20 and 427  $\mu$ g/l respectively, whereas the median and maximum AMPA concentrations were 0.43 and 397  $\mu$ g/l, respectively (Table 1). The median %AMPA ratio (in 316 samples) was 63% with an interquartile range of 29-85% (Figure 8).

A total of 104 lake, pond, or wetland samples were collected from 65 sites in California, Colorado, District of Columbia, Florida, Iowa, Kansas, Maryland, Maine, Michigan, Minnesota, Nebraska, New Jersey, Oregon, South Dakota, Vermont, and Wyoming. Glyphosate and AMPA were detected at least once in samples from 27 sites. Glyphosate was detected in 33.7% and AMPA in 29.8% of lake, pond, or wetland samples (Figures 3 and 7). The median and maximum glyphosate concentrations in these samples

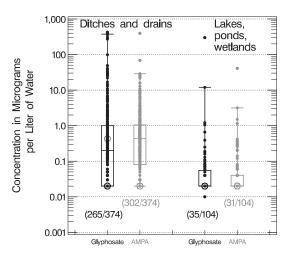


FIGURE 7. Boxplot-Dot Plots Showing Concentrations of Glyphosate and AMPA for Ditch and Drain Samples and for Lake, Pond, and Wetland Samples (numbers in parentheses are number of detections/number of samples, open circle is reporting level).

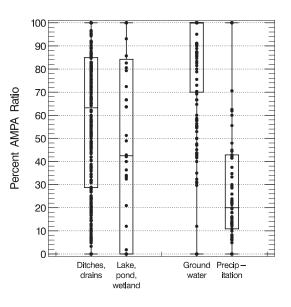


FIGURE 8. Boxplot-Dot Plots Showing the Percent AMPA Ratio for Ditches and Drains; Lake, Pond, and Wetland; Groundwater; and Precipitation Samples.

were <0.02 and 301  $\mu$ g/l, respectively, whereas the median and maximum AMPA concentrations were <0.02 and 41  $\mu$ g/l, respectively. The median %AMPA ratio (in 44 samples) was 42% with an interquartile range of 0-84% (Figure 8).

### Groundwater and Precipitation

A total of 1,171 groundwater samples were collected from 807 sites in California, Delaware, Florida, Georgia, Iowa, Idaho, Illinois, Indiana, Kansas, Mary-

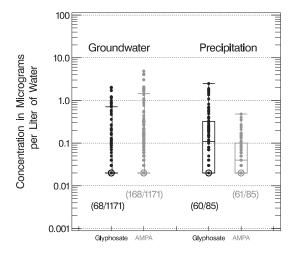


FIGURE 9. Boxplot-Dot Plots Showing Concentrations of Glyphosate and AMPA for Groundwater and Precipitation Samples (numbers in parentheses are number of detections/number of samples, open circle is reporting level).

land, Maine, Michigan, Mississippi, North Carolina, New Jersey, New York, Ohio, Oregon, South Carolina, Texas, Washington, Wisconsin, and Wyoming. Glyphosate was detected at least once in samples from 32 sites, whereas AMPA was detected at least once at 57 sites. Glyphosate was detected in 5.8% and AMPA in 14.3% of groundwater samples (Figures 3 and 9). The median and maximum glyphosate concentrations in these samples were <0.02 and 2.03 µg/l, respectively, whereas the median and maximum AMPA concentrations were <0.02 and 4.88 µg/l, respectively (Table 1). The median %AMPA ratio (in 179 samples) was 100% with an interquartile range of 70-100% (Figure 8).

A total of 85 precipitation samples were collected from three sites in Iowa, Indiana, and Mississippi. Glyphosate and AMPA were detected at least once in samples from all three sites. Glyphosate was detected in 70.6% and AMPA in 71.8% of precipitation samples (Figures 3 and 9). The median and maximum glyphosate concentrations in precipitation samples were 0.11 and 2.50 µg/l, respectively, whereas the median and maximum AMPA concentrations were 0.04 and 0.48 µg/l, respectively (Table 1). The median %AMPA ratio (in 69 samples) was 20% with an interquartile range of 11-43% (Figure 8).

#### Temporal Patterns

Most of the samples analyzed in this study were not collected with the intention of identifying temporal patterns or trends. No sites had results from all years and most sites only had results from one or two years. A change in the laboratory reporting level in 2004 also complicates the interpretation of temporal

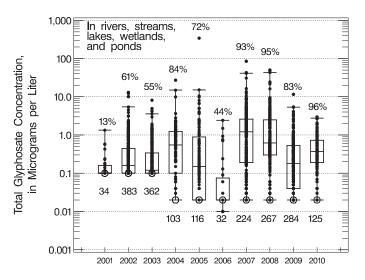


FIGURE 10. Boxplot-Dot Plots Showing Number of Samples (number below boxplots), Percentage Detections (number above boxplots), and Total Glyphosate Concentrations, by Year 2001-2010, for Surface Water Samples from Rivers, Streams, Lakes, Wetlands, and Ponds (open circle is reporting level).

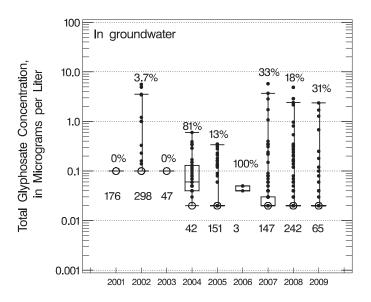


FIGURE 11. Boxplot-Dot Plots Showing Number of Samples (number below boxplots), Percentage Detections (number above boxplots), and Total Glyphosate Concentrations, by Year 2001-2009, for Groundwater Samples (open circle is reporting level).

patterns of glyphosate and AMPA occurrence. A plot of the total glyphosate concentration (sum of glyphosate and AMPA) by year for all surface water samples (Figure 10) provides limited indication of increases in detection frequency and median concentration. A plot of the total glyphosate concentration by year for all groundwater samples (Figure 11) provides no indication of increases in detection frequency or concentration. At six stream sites and three river sites, there were multiple samples from multiple years, both early (prior to 2006) and late (2006 and later) during the period of study. Streamflow data were acquired for these sites and used to calculate estimates of instantaneous daily total glyphosate flux on dates when samples were collected (Figure 12).

The Wilcoxon signed-rank test was used to gain a measure of the statistical significance of differences in streamflows, total glyphosate concentrations, and instantaneous daily total glyphosate fluxes between the early period (2001-2005) and the later period (2006-2010). Results (Table 2) indicated that streamflow was significantly (p < 0.05) larger for the late period samples at two sites; significantly smaller for the late period at two sites; larger, but not statistically significantly larger at three sites; and smaller, but not significantly smaller at two sites. Hence, there was a mix of changing streamflow conditions at the nine sites. In contrast, total glyphosate concentrations were significantly larger for the late period samples at five sites, and larger, but not significantly larger at the other four sites. Instantaneous daily total glyphosate fluxes were significantly larger for the late period samples at four sites, larger, but not significantly larger at three sites; and smaller, but

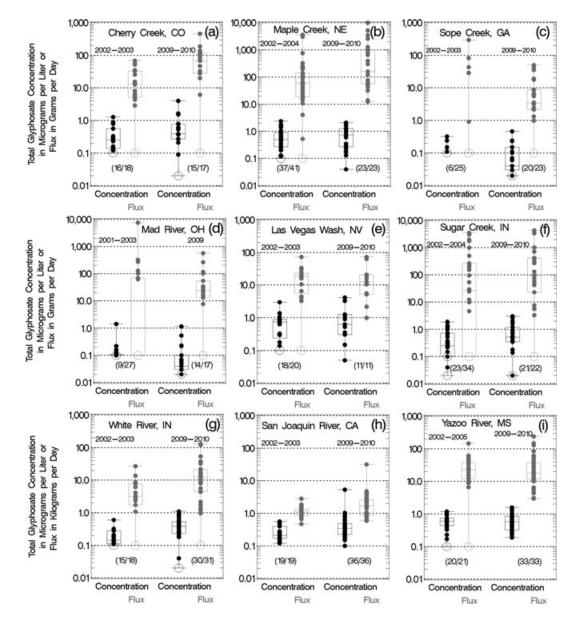


FIGURE 12. Boxplot-Dot Plots Showing Total Glyphosate Concentrations and Instantaneous Daily Total Glyphosate Fluxes for Early and Late Samples from (a) Cherry Creek, Colorado; (b) Maple Creek, Nebraska; (c) Sope Creek, Georgia; (d) Mad River, Ohio; (e) Las Vegas Wash, Nevada; (f) Sugar Creek, Indiana; (g) White River, Indiana; (h) San Joaquin River, California; and (i) Yazoo River, Mississippi (numbers in parentheses are number of detections/number of samples, open circle is reporting level).

|                               | Streamflow |                      | Concentration |                      | Instantaneous Flux |                      |
|-------------------------------|------------|----------------------|---------------|----------------------|--------------------|----------------------|
| Site                          | p Value    | 2006-2010 Values Are | p Value       | 2006-2010 Values Are | p Value            | 2006-2010 Values Are |
| Cherry Creek, Colorado        | < 0.001    | Larger               | 0.08          | Larger               | 0.003              | Larger               |
| Maple Creek, Nebraska         | < 0.001    | Larger               | 0.206         | Larger               | 0.002              | Larger               |
| Sope Creek, Georgia           | 0.227      | Larger               | 0.001         | Larger               | 0.001              | Larger               |
| Mad River, Ohio               | 0.003      | Smaller              | 0.039         | Larger               | 0.078              | Larger               |
| Las Vegas Wash, Nevada        | < 0.001    | Smaller              | 0.591         | Larger               | 0.698              | Smaller              |
| Sugar Creek, Indiana          | 0.731      | Smaller              | 0.017         | Larger               | 0.142              | Larger               |
| White River, Indiana          | 0.583      | Larger               | 0.001         | Larger               | 0.003              | Larger               |
| San Joaquin River, California | 1.0        | Larger               | 0.034         | Larger               | 0.074              | Larger               |
| Yazoo River, Mississippi      | 0.729      | Smaller              | 0.936         | Larger               | 0.887              | Smaller              |

TABLE 2. Wilcoxon Signed-Rank Test *p*-Values and Direction of the Differences between Early (2001-2005) andLate (2006-2010) Values of Streamflow, Total Glyphosate Concentration, and Instantaneous Daily Total Glyphosate Flux<br/>at Nine Sites (bold values indicate that differences are statistically significant at the  $p \leq 0.05$  level).

not significantly smaller at two sites, both of which have smaller streamflow in the later period (Table 2).

### DISCUSSION

Results described here indicate that glyphosate and AMPA are mobile and occur widely in the environment. It was uncommon for glyphosate to be detected without AMPA, happening in only 2.3% of all samples, whereas AMPA was detected without glyphosate in 17.9% of all samples. Glyphosate was detected in 52.5% of stream and 53.1% of large river samples, whereas AMPA was detected in 71.6% of stream and 89.3% of large river samples (Table 1, Figure 3). Glyphosate and AMPA were detected in very large rivers such as the Mississippi with drainage areas in the millions of square kilometers and in headwaters streams with drainage areas less than 10 km<sup>2</sup>. These detection frequencies are greater than those determined from samples collected in 2007 from urban and rural streams in Ontario, 33 and 32%, respectively (Byer et al., 2008). The detection frequencies also are much greater than those identified by Struger et al. (2008) in Ontario streams, 21% for glyphosate and 3% for AMPA in 502 samples, however, the analytical reporting level for the method used in that study were substantially higher (5  $\mu$ g/l for glyphosate and 20 µg/l for AMPA) than the reporting levels used in this study, emphasizing the importance of low reporting levels for targeted analytes and their degradates in environmental occurrence studies.

Most observed concentrations of glyphosate were well below existing health benchmarks and levels of concern for humans or wildlife, and none exceeded the U.S. Environmental Protection Agency's Maximum Contaminant Level of 700  $\mu$ g/l or Canadian

short-term (27,000 µg/l) and long-term (800 µg/l) freshwater aquatic life standards (Canadian Council of Ministers of the Environment, 2012). Median glyphosate concentrations in all hydrologic settings (other than sediment and precipitation) were less than or equal to 0.2 µg/l and median AMPA concentration in all hydrologic settings (other than sediment) were less than or equal to 0.45  $\mu$ g/l. In isolated samples glyphosate concentrations in surface water approached a level (about 400  $\mu$ g/l) that could be of concern for the survival of some amphibian species (King and Wagner, 2010), but only if the surfactants and other adjuvants used in glyphosate formulations were also present. While concentrations of glyphosate and AMPA were below the levels of concern for humans or wildlife, pesticides (and other environmental contaminants) are often detected in mixtures, and the ecosystem effects of chronic low-level exposures to pesticide mixtures are uncertain. Hence, the environmental health risk of these low-level detections of glyphosate, AMPA, and the potential associated adjuvants and mixtures remains to be determined.

One likely reason for the high detection frequencies is simply the widespread and increasing use of products containing glyphosate in the U.S. The extensive use of tile/subsurface drains in many agricultural regions in the U.S. is another factor that could contribute to the higher than expected frequency of detection of glyphosate and AMPA in U.S. streams and rivers. Others have suggested that glyphosate transport via tile drains could be significant (Stone and Wilson, 2006), and both glyphosate and AMPA were detected frequently and often at elevated concentrations in ditch and drain samples analyzed in this study (Figures 3 and 7). The widespread use of glyphosate for nonagricultural purposes and the frequent occurrence of glyphosate and AMPA in precipitation, and the discharge of AMPA by WWTP and septic tanks also could contribute to frequent detections in surface waters.

Another potentially important reason why glyphosate occurrence is more common than would be expected is that surfactants and other adjuvants are almost always included with glyphosate in commercial products, or added as "tank-mixtures" just prior to application. Several studies have demonstrated that the mobility of typically immobile pesticides can be increased in the presence of surfactants (Grant et al., 2011). Surfactants can increase the apparent water solubility of a pesticide, influence biodegradation, and effect soil structure and related adsorption and desorption processes (Katagi, 2008). The type and concentration of the surfactant is important and there is typically a critical concentration at which surfactant micelles form. When below this level, surfactants may act to increase the sorption of a pesticide to soils, whereas when above this level they would decrease the sorption to soils and increase mobility (Haigh, 1996).

The detection of glyphosate in 5.8% and AMPA in 14.3% of groundwater samples (Figures 3 and 9) was about what was expected. While most prior reviews of the occurrence or expected occurrence of glyphosate and AMPA suggested that both compounds were "unlikely to leach into groundwater" due to their strong adsorptive characteristics (U.S. Environmental Protection Agency, 1993; Giesy et al., 2000; Cerdeira and Duke, 2006; Borggaard and Gimsing, 2008), at least one (Vereecken, 2005) suggested some potential for movement after heavy rainfall in the presence of preferential flow paths. Also, one recent study (Sanchis et al., 2011), which used methods that had comparably low analytical limits of quantification (~10 ng/l), found glyphosate in 41% of groundwater samples from Catalonia, Spain. The detection frequencies for glyphosate and AMPA in this study, which includes shallow and deep wells, and wells from nonagricultural areas, are similar to those determined for other high use herbicides such as acetochlor, atrazine, alachlor, metolachlor, and their degradates in Iowa groundwater (Kolpin et al., 2000). One prior study (U.S. Environmental Protection Agency, 2002) had detected glyphosate in less than 0.1% of groundwater systems used as drinking water supplies, however, the samples were collected between 1992 and 1997, well before the rapid increase in glyphosate use, and the analytical reporting level for glyphosate was 6  $\mu$ g/l.

The detection of glyphosate and AMPA in more than 70% of the precipitation samples (Figure 3) was not expected due to their low vapor pressures and strong adsorptive characteristics, however, spray drift from such a heavily used pesticide is always possible (Giesy *et al.*, 2000). Other herbicides with similar use patterns such as atrazine and metolachlor also commonly occur in precipitation in agricultural areas (Goolsby *et al.*, 1997; Vogel *et al.*, 2008; Schummer *et al.*, 2010). A more detailed analysis of the occurrence of glyphosate and AMPA in these precipitation samples and associated air samples was recently provided by Chang *et al.* (2011), who indicated that both spray drift and wind erosion are important sources of glyphosate to the atmosphere and that precipitation is very effective at removing glyphosate and AMPA from the atmosphere.

The %AMPA values provide some information on the source, fate, and transport of glyphosate in the environment with lower values suggesting recent or proximal input of glyphosate and higher values suggesting more residence time or distance between input and the measured occurrence. More than 75% of %AMPA values from sediment, soil water, large river, stream, and groundwater samples (Figures 5 and 8) were greater than 50 indicating that AMPA tends to occur at higher concentrations than glyphosate in these environmental settings. Coupe et al. (2012) suggest that the timing of rainfall runoff events relative to glyphosate and the amount of glyphosate and AMPA in the soil reservoir from previous applications controls %AMPA values in surface water. Larger %AMPA values are expected when rainfall runoff events occur later in the season or when there is a larger reservoir of available AMPA than glyphosate in the soil reservoir or when there is sufficient travel distance/residence time between source applications and transport to surface water to allow for the degradation of glyphosate to AMPA. Coupe et al. (2012) also suggest that the %AMPA values should increase with increases in drainage area. In this study, %AMPA values from ditch and drain; and lake, pond, and wetland samples both ranged from 0 to 100% with median values of 63 and 42%, respectively. This result suggests that these site types span a wide range of hydrologic conditions, but that more often than with streams or rivers, they are closely connected to the source applications (in time or space). Groundwater samples had the highest %AMPA values (Figure 8) with a median value of 100 indicating that water in this hydrologic setting is the farthest (in residence time or space) from the source application. Greater sorption of glyphosate relative to AMPA in soils also may contribute to the higher %AMPA values in groundwater samples. More than 75% of %AMPA values from precipitation samples (Figure 8) were less than 50 indicating that glyphosate tends to occur at higher concentrations than does AMPA in this environmental setting.

Although most of the samples analyzed in this study were not collected with the intention of identifying temporal patterns or trends, six stream sites and three river sites had multiple samples from multiple years, both early (prior to 2006) and late (2006 and later) during the period of study. Total glyphosate concentrations were larger in 2006-2010 than in 2001-2005 at all nine sites and those differences are statistically significant (p < 0.05) at five of the sites. And while streamflow was larger in 2006-2010 at five sites and smaller in 2006-2010 at four sites, instantaneous daily flux values were larger in 2006-2010 at seven of nine sites, and four sites indicate statistically significant increases, and no sites indicate statistically significant decreases (Figure 12, Table 2).

### CONCLUSIONS

This investigation is the largest and most comprehensive assessment of the environmental occurrence of glyphosate and AMPA in the U.S. conducted to date, summarizing the results of 3,732 environmental water and sediment samples and 1,018 quality assurance samples collected between 2001 and 2010 from 38 states and the District of Columbia. The results indicate that glyphosate and AMPA are mobile, occur widely in the environment, and have both agricultural and urban sources. It was uncommon for glyphosate to be detected without AMPA, occurring in only 2.3% of all samples, whereas AMPA was detected without glyphosate in 17.9% of all samples. Glyphosate and AMPA occurred widely in surface water with one or both compounds being detected at least once at 59% of 470 sites. Glyphosate and AMPA were detected with similar frequency in large rivers such as the Mississippi with drainage areas equal to or greater than 10,000 km<sup>2</sup> and in smaller streams with drainage areas less than 10,000 km<sup>2</sup>. Glyphosate and AMPA occurred less widely in groundwater or soil water with one or both compounds being detected at least once at 8.4% of 820 sites. Glyphosate was detected in more than 50% of soil and sediment samples, and water samples from ditches and drains, precipitation, large rivers, and streams. Glyphosate was detected in less than 40% of water samples from lakes, ponds, and wetlands; soil water; and groundwater. AMPA was detected in more than 50% of soil and sediment samples, and water samples from large rivers, ditches and drains, precipitation, streams, and soil water. AMPA was detected in less than 30% of water samples from lakes, ponds, and wetlands; and groundwater. AMPA was detected more frequently than glyphosate in all hydrologic settings except lakes, ponds, and wetlands. These differences in detection frequencies for glyphosate and AMPA are likely due to differences in source proximity, water travel time, water residence time, degradation processes, and other natural processes.

The %AMPA values confirm that AMPA is detected at higher concentrations than glyphosate in most

hydrologic settings, with groundwater and soil water samples having the highest values; and precipitation and lake, pond, and wetland samples having the lowest values. These results indicate that the glyphosate in precipitation and wetland, pond, and lake water samples was more closely associated with source applications or has had less opportunity to degrade than did the glyphosate occurring in the other hydrologic settings, and that glyphosate reaching soil water and groundwater had the most opportunity to degrade. Median glyphosate concentrations in all hydrologic settings (other than sediment) were less than or equal to 0.2 µg/l and median AMPA concentration in all hydrologic settings (other than sediment) were less than or equal to  $0.45 \,\mu g/l$ , emphasizing the importance of low limits of detection for targeted analytes and their degradates in environmental occurrence studies.

Although most samples analyzed in this study were not collected with the intention of identifying temporal patterns or trends, results from nine surface water sites that had multiple samples from both the early (2001-2005) and late (2006-2010) study periods provide an indication of increases in glyphosate and AMPA detection frequency, median concentrations, and instantaneous daily fluxes. Finally, the results indicate that glyphosate and AMPA frequently add to the chronic low-level exposures to mixtures of pesticides and pesticide degradation products that plants and animals experience in a wide range of ecosystems in the U.S.

### ACKNOWLEDGMENTS

The authors would like to thank all of the USGS Water Science Center personnel who collected samples, processed and managed data, or otherwise contributed to the creation of this large data set including Donald Goolsby, Dana Kolpin, Mike Thurman, Mike Meyer, Betty Scribner, Steve Kalkhoff, Jeff Frey, Richard Coupe, Paul Capel, and Robert Gilliom. The analytical skills, technical support, and data management skills of the staff at the USGS Organic Geochemistry Research Laboratory in Lawrence, Kansas, were essential in completion of this manuscript. Helpful comments from Michael Focazio, Dana Kolpin, and two anonymous reviewers greatly improved the manuscript. The authors gratefully acknowledge the financial support from the USGS Toxics Substances Hydrology Program and the USGS Amphibian Research and Monitoring Initiative (ARMI). Any use of trade, firm, or product names is for descriptive purposes only and does not imply endorsement by the U.S. Government.

### LITERATURE CITED

Aspelin, A.L., 1997. Pesticide Industry Sales and Usage: 1994 and 1995 Market Estimates. U.S. Environmental Protection Agency, Office of Pesticide Programs. EPA-733-R-97-002, 35 pp.

- Baker, N.T., W.W. Stone, J.T. Wilson, and M.T. Meyer, 2006. Occurrence and Transport of Agricultural Chemicals in Leary Weber Ditch Basin, Hancock County, Indiana, 2003-04. U.S. Geological Survey Scientific Investigations Report 2006-5251, 44 pp.
- Battaglin, W.A., D.W. Kolpin, E.A. Scribner, K.M. Kuivila, and M.W. Sandstrom, 2005. Glyphosate, Other Herbicides, and Transformation Products in Midwestern Streams, 2002. Journal of the American Water Resources Association 41(2):323-332.
- Battaglin, W.A., K.C. Rice, M.J. Focazio, S. Salmons, and R.X. Barry, 2009. The Occurrence of Glyphosate, Atrazine, and Other Pesticides in Vernal Pools and Adjacent Streams in Washington, DC, Maryland, Iowa, and Wyoming, 2005-2006. Environmental Monitoring and Assessment 155:281-307.
- Benachour, N. and G.E. Seralini, 2009. Glyphosate Formulations Induce Apoptosis and Necrosis in Human Umbilical, Embryonic, and Placental Cells. Chemical Research in Toxicology 22:97-105.
- Benachour, N., H. Sipahutar, S. Moslemi, C. Gasnier, C. Travert, and G. Seralini, 2007. Time- and Dose-Dependent Effects of Roundup<sup>®</sup> on Human Embryonic and Placental Cells. Archives of Environmental Contamination and Toxicology 53:126-133.
- Benbrook, C.M., 2012. Impacts of Genetically Engineered Crops on Pesticide Use in the U.S.—the First Sixteen Years. Environmental Sciences. Europe 24:24, doi: 10.1186/2190-4715-24-24
- Bergstrom, L., E. Borjesson, and J. Stenstrom, 2011. Laboratory and Lysimeter Studies of Glyphosate and Aminomethylphosphonic Acid in a Sand and a Clay Soil. Journal of Environmental Quality 40:98-108.
- Blanchoud, H., E. Moreau-Guigon, F. Farrugia, M. Chevreuil, and J.M. Mouchel, 2007. Contribution by Urban and Agricultural Pesticide Uses to Water Contamination at the Scale of the Marne Watershed. Science of the Total Environment 375:168-179.
- Borggaard, O.K. and A.L. Gimsing, 2008. Fate of Glyphosate in Soil and the Possibility of Leaching to Ground and Surface Waters: A Review. Pest Management Science 64:441-456.
- Botta, F., G. Lavison, G. Couturier, F. Alliot, E. Moreau-Guigon, N. Fachon, B. Guery, M. Chevreuil, and H. Blanchoud, 2009. Transfer of Glyphosate and Its Degradate AMPA to Surface Waters through Urban Sewerage Systems. Chemosphere 77:133-139.
- Brausch, J.M., B. Beall, and P.N. Smith, 2007. Acute and Sub-Lethal Toxicity of Three POEA Surfactant Formulations to Daphnia Magna. Bulletin of Environmental Contamination and Toxicology 78:510-514.
- Bringolf, R.B., W.G. Cope, S. Mosher, M.C. Barnhart, and D. Shea, 2007. Acute and Chronic Toxicity of Glyphosate Compounds to Glochidia and Juveniles of Lampsilis Siliquoidea (Unionidae). Environmental Toxicology and Chemistry 26(10):2094-2100.
- Busse, M.D., A.W. Ratcliff, C.J. Stestak, and R.F. Powers, 2010. Glyphosate Toxicity and the Effects of Long-Term Vegetation Control on Soil Microbial Communities. Soil Biology & Biochemistry 33:1777-1789.
- Byer, J.D., J. Struger, P. Klawunn, A. Todd, and E. Sverko, 2008. Low Cost Monitoring of Glyphosate in Surface Waters Using the ELISA Method: An Evaluation. Environmental Science and Technology 42:6052-6057.
- Canadian Council of Ministers of the Environment, 2012. Scientific Criteria Document for the Development of the Canadian Water Quality Guidelines for the Protection of Aquatic Life: Glyphosate. PN 1469, ISBN 978-1-896997-83-4 PDF, 68 pp.
- Cauble, K. and R.S. Wagner, 2005. Sublethal Effects of Herbicide Glyphosate on Amphibian Metamorphosis and Development. Bulletin of Environmental Contamination and Toxicology 75:429-435.
- Cerdeira, A.L. and S.O. Duke, 2006. The Current Status and Environmental Impacts of Glyphosate-Resistant Crops: A Review. Journal of Environmental Quality 35:1633-1658.

- Chang, F., M.F. Simcik, and P.D. Capel, 2011. Occurrence and Fate of the Herbicide Glyphosate and Its Degradate Aminomethylphosphonic Acid in the Atmosphere. Environmental Toxicology and Chemistry 30(3):548-555.
- Coupe, R.H., S.J. Kalkhoff, P.D. Capel, and C. Gregoire, 2012. Fate and Transport of Glyphosate and Aminomethylphosphonic Acid in Surface Waters of Agricultural Basins. Pest Management Science 68(1):16-30.
- Dallegrave, E., F.D. Mantese, R.S. Coelho, J.D. Pereira, P.R. Dalsenter, and A. Langeloh, 2003. The Teratogenic Potential of the Herbicide Glyphosate-Roundup in Wistar Rats. Toxicology Letters 142:45-52.
- Dallegrave, E., F.D. Mantese, R.T. Oliveira, A.J.M. Andrade, P.R. Dalsenter, and A. Langeloh, 2007. Pre- and Postnatal Toxicity of the Commercial Glyphosate Formulation in Wistar Rats. Archives of Toxicology 81(9):665-673.
- Daruich, J., F. Zirulnik, and M.S. Gimenez, 2001. Effect of the Herbicide Glyphosate on Enzymatic Activity in Pregnant Rats and Their Foetuses. Environmental Research 85(3):226-231.
- Dill, G.M., C.A. CaJacob, and S.R. Padgette, 2008. Current and Future Glyphosate-Resistant Crops: Adoption, Use and Future Considerations. Pest Management Science 64(4):326-331.
- Dinehart, S.K., L.M. Smith, S.T. McMurry, T.A. Anderson, P.N. Smith, and D.A. Haukos, 2009. Toxicity of a Glufosinate and Several Glyphosate-Based Herbicides to Juvenile Amphibians from the Southern High Plains, USA. Science of the Total Environment 407:1065-1071.
- Donaldson, D., T. Kiely, and A. Grube, 2002. Pesticide Industry Sales and Usage: 1998 and 1999 Market Estimates. U.S. Environmental Protection Agency, Office of Pesticide Programs. EPA-733-R-02-001, 33 pp.
- Duke, S.O. and S.B. Powles, 2008. Mini-Review Glyphosate: A Oncein-a-Century Herbicide. Pest Management Science 64:319-325.
- Edginton, A.N., P.M. Sheridan, G.R. Stephenson, D.G. Thompson, and H.J. Boermans, 2004. Comparative Effects of pH and Vision<sup>®</sup> on Two Life Stages of Four Anuran Amphibian Species. Environmental Toxicology and Chemistry 23(4):815-822.
- FAO, 1997. Pesticide Residues in Food-1997-Aminomethylphosphonic Acid (AMPA). FAO Panel of Experts on Pesticides Residues and the WHO Core Assessment Group. http://www.inchem.org/documents/jmpr/jmpmono/v097pr04.htm, accessed January 2013.
- Franz, J.E., M.K. Mao, and J.A. Sikorski, 1997. Glyphosate: A Unique Global Herbicide. ACS Monograph 189, American Chemical Society, Washington, D.C., pp. 163-175.
- Gahl, M., B.D. Pauli, and J.E. Houlahan, 2011. Effects of Chytrid Fungus and a Glyphosate-Based Herbicide on Survival and Growth of Wood Frogs (*Lithobates sylvaticus*). Ecological Applications 21(7):2521-2529.
- Gianessi, L., 2008. Economic Impacts of Glyphosate-Resistant Crops. Pest Management Science 64(4):346-352.
- Gianessi, L. and N. Reigner, 2006. Pesticide Use in US Crop Production 2002—with Comparison to 1992 and 1997. CropLife Foundation, Washington, D.C.
- Gianessi, L. and S. Sankula, 2003. The Value of Herbicides in U.S. Crop Production. National Center for Food & Agricultural Policy, Washington, D.C. http://croplifefoundation.files.wordpress. com/2012/07/herbicide-benefits-2003-full.pdf, accessed December 2012.
- Giesy, J.P., S. Dobson, and K. Solomon, 2000. Ecotoxicological Risk Assessment for Roundup<sup>®</sup> Herbicide. Reviews of Environmental Contaminant Toxicology 167:35-120.
- Gilliom, R.J., J.E. Barbash, C.G. Crawford, P.A. Hamilton, J.D. Martin, N. Nakagaki, L.H. Nowel, J.C. Scott, P.E. Stackelberg, G.P. Thelin, and D.M. Wolock, 2006. Pesticides in the Nation's Streams and Ground Water, 1992-2001. U.S. Geological Survey Circular 1291, 173 pp.

- Goolsby, D.A., E.M. Thurman, M.L. Pomes, M.T. Meyer, and W.A. Battaglin, 1997. Herbicides and Their Metabolites in Rainfall: Origin, Transport, and Deposition Patterns Across the Midwestern and Northeastern United States, 1990-1991. Environmental Science and Technology 31(5):1325-1333.
- Grant, S., M. Mortimer, G. Stevenson, D. Malcolm, and C. Gaus, 2011. Facilitated Transport of Dioxins in Soil Following Unintentional Release of Pesticide-Surfactant Formulations. Environmental Science and Technology 45:406-411.
- Griesinger, L.M., S.C. Evans, and A.L. Rypstra, 2011. Effects of a Glyphosate-Based Herbicide on Mate Location in a Wolf Spider That Inhabits Agroecosystems. Chemosphere 84:1461-1466.
- Grube, A., D. Donaldson, T. Kiely, and L. Wu, 2011. Pesticide Industry Sales and Usage: 2006 and 2007 Market Estimates. U.S. Environmental Protection Agency, Biological and Economic Analysis Division, Office of Pesticide Programs, Office of Prevention, Pesticides, and Toxic Substances, 33 pp.
- Grunewald, K., W. Schmidt, C. Unger, and G. Hanschmann, 2001. Behavior of Glyphosate and Aminomethylphosphonic Acid (AMPA) in Soils and Water of Reservoir Radeburg II Catchment (Saxony/Germany). Journal of Plant Nutrition and Soil Science 164:65-70.
- Haigh, S.D., 1996. A Review of the Interaction of Surfactants with Organic Contaminants in Soil. Science of the Total Environment 185:161-170.
- Haney, R.L., S.A. Senseman, F.M. Hons, and D.A. Zuberer, 2000. Effect of Glyphosate on Soil Microbial Activity and Biomass. Weed Science 48:89-93.
- Hanke, I., I. Wittmer, S. Bischofberger, C. Stamm, and H. Singer, 2010. Relevance of Urban Glyphosate Use for Surface Water Quality. Chemosphere 81:422-429.
- Helsel, D.R. and R.M. Hirsch, 2002. Statistical Methods in Water Resources. Techniques of Water-Resources Investigations of the United States Geological Survey, book 4, Hydrologic Analysis and Interpretation, chap. A3, 524 pp. http://www.practicalstats. com/aes/aesbook/files/HelselHirsch.PDF, accessed May 2008.
- HERA, 2004. Human and Environmental Risk Assessment on Ingredients of European Household Cleaning Products—Phosphonates, 114 pp.
- Horowitz, J., R. Ebel, and K. Ueda, 2010. "No-Till" Farming Is a Growing Practice. United States Department of Agriculture ERS, Economic Information Bulletin No. 70, 22 pp.
- Howe, C.M., M. Berrill, B.D. Pauli, C.C. Helbing, K. Werry, and N. Veldhoen, 2004. Toxicity of Glyphosate-Based Pesticides to Four North American Frog Species. Environmental Toxicology and Chemistry 23(8):1928-1934.
- Jones, D.K., J.I. Hammond, and R.A. Relyea, 2010. Roundup and Amphibians: The Importance of Concentration, Application Time, and Stratification. Environmental Toxicology and Chemistry 29(9):2016-2025.
- Katagi, T., 2008. Surfactant Effects on Environmental Behaviour of Pesticides. *In*: Reviews of Environmental Contamination and Toxicology, D.M. Whitacre (Editor). Springer, New York, pp. 71-177.
- Kelly, D.W., R. Poulin, D.M. Tompkins, and C.R. Townsend, 2010. Synergistic Effects of Glyphosate Formulation and Parasite Infection on Fish Malformations and Survival. Journal of Applied Ecology 47(2):498-504.
- Kiely, T., D. Donaldson, and A. Grube, 2004. Pesticide Industry Sales and Usage: 2000 and 2001 Market Estimates. U.S. Environmental Protection Agency, Office of Prevention, Pesticides, and Toxic Substances (7503C) EPA-733-R-04-001, 34 pp.
- King, J.J. and R.S. Wagner, 2010. Toxic Effects of the Herbicide Roundup Regular on Pacific Northwestern Amphibians. Northwestern Naturalist 91:318-324.
- Kolpin, D.W., D.J. Schnoebelen, and E.M. Thurman, 2004. Degradates Provide Insight to Spatial and Temporal Trends of Herbicides in Ground Water. Groundwater 42(4):601-608.

- Kolpin, D.W., E.M. Thurman, E.A. Lee, M.T. Meyer, E.T. Furlong, and S.T. Glassmeyer, 2006. Urban Contributions of Glyphosate and Its Degradate AMPA to Streams in the United States. Science of the Total Environment 354(2-3):191-197.
- Kolpin, D.W., E.M. Thurman, and S.M. Linhart, 2000. Finding Minimal Herbicide Concentrations in Ground Water? Try Looking for Their Degradates. Science of the Total Environment 248(2-3):115-122.
- Lajmanovich, R.C., M.T. Sandoval, and P.M. Peltzer, 2003. Induction of Mortality and Malformation in Scinax Nasicus Tadpoles Exposed to Glyphosate Formulations. Bulletin of Environmental Contamination and Toxicology 70:612-618.
- Lee, E.A., L.R. Zimmerman, B.S. Bhullar, and E.M. Thurman, 2002. Linker-Assisted Immunoassay and Liquid Chromatography/Mass Spectrometry for the Analysis of Glyphosate. Analytical Chemistry 74:4937-4943.
- Lenkowski, J.R., G. Sanchez-Bravo, and K.A. McLaughlin, 2010. Low Concentrations of Atrazine, Glyphosate, 2,4-Dichlorophenoxyacetic Acid, and Triadimefon Exposure Have Diverse Effects on *Xenopus laevis* Organ Morphogenesis. Journal of Environmental Science 22(9):1305-1308.
- Loos, R., G. Locoro, S. Comero, S. Contini, D. Schwesig, F. Werres, P. Balsaa, O. Gans, S. Weiss, L. Blaha, M. Bolchi, and B.M. Gawlik, 2010. Pan-European Survey on the Occurrence of Selected Polar Organic Persistent Pollutants in Ground Water. Water Research 44:4115-4126.
- Mañas, F., L. Peralta, J. Raviolo, O. Ovando, A. Weyers, L. Ugina, C. Gonzalez, I. Larripa, and N. Gorla, 2009. Genotoxicity of AMPA, the Environmental Metabolite of Glyphosate, Accessed by the Comet Assay and Cytogenetic Tests. Ecotoxicology and Environmental Safety 72:834-837.
- Mann, R.M., R.V. Hyne, C.B. Choung, and S.P. Wilson, 2009. Amphibians and Agricultural Chemicals: Review of the Risks in a Complex Environment. Environmental Pollution 157:2903-2927.
- McCarthy, K.A., D.C. Lampe, and P.D. Capel, 2011. Discrete and Continuous Water-Quality Data and Hydrologic Parameters from Seven Agricultural Watersheds in the United States, 2002-09. U.S. Geological Survey Data Series 603, 10 pp.
- Mesnage, R., B. Bernay, and G.E. Seralini, 2012. Ethoxylated Adjuvants of Glyphosate-Based Herbicides Are Active Principles of Human Cell Toxicity. Toxicology 313(2-3):122-128.
- Meyer, M.T., K.A. Loftin, E.A. Lee, G.H. Hinshaw, J.D. Dietze, and E.A. Scribner, 2009. Determination of Glyphosate, Its Degradation Product Aminomethlyphosphonic Acid, and Glufosinate, in Water by Isotope Dilution and Online Solid-Phase Extraction and Liquid Chromatography/Tandem Mass Spectrometry. U.S. Geological Survey Techniques and Methods, Book 5, Chap. A10, 32 pp.
- Monsanto, 2009. Backgrounder: History of Monsanto's Glyphosate Herbicides. http://www.monsanto.com/monsanto/content/products/ productivity/roundup/back\_history.pdf, *accessed* January 2009.
- Moore, L.J., L. Fuentes, J.H. Rodgers, W.W. Bowerman, G.K. Yarrow, W.Y. Chao, and W.C. Bridges, 2012. Relative Toxicity of the Components of the Original Formulation of Roundup<sup>®</sup> to Five North American Anurans. Ecotoxicology and Environmental Safety 78:128-133.
- National Pesticide Information Center, 2008. Glyphosate Technical Fact Sheet. Oregon State University, National Pesticide Information Center, Corvallis, Oregon, 14 pp.
- Nowack, B., 2003. Environmental Chemistry of Phosphonates. Water Research 37:2533-2546.
- Owens, M.D.K., 2008. Evolved Glyphosate-Resistant Weeds and Weed Shifts: Weed Species Shifts in Glyphosate-Resistant Crops. Pest Management Science 64(4):377-387.
- Paganelli, A., V. Gnazzo, H. Acosta, S.L. Lopez, and A.E. Carrasco, 2010. Glyphosate-Based Herbicides Produce Teratogenic Effects

on Vertebrates by Impairing Retinoic Acid Signalling. Chemical Research in Toxicology 23(10):1586-1595.

- Pesce, S., I. Batisson, C. Bardot, C. Fajon, C. Portelli, B. Montuelle, and J. Bohatier, 2009. Response of Spring and Summer Riverine Microbial Communities Following Glyphosate Exposure. Ecotoxicology and Environmental Safety 72:1905-1912.
- Powles, S.B., 2008. Evolved Glyphosate-Resistant Weeds Around the World: Lessons to be Learnt. Pest Management Science 64(4):360-365.
- Relyea, R.A., 2005a. The Lethal Impacts of Roundup<sup>®</sup> and Predatory Stress on Six Species of North American Tadpoles. Archives of Environmental Contaminant Toxicology 48:351-357.
- Relyea, R.A., 2005b. The Lethal Impact of Roundup<sup>®</sup> on Aquatic and Terrestrial Amphibians. Ecological Applications 15(4):1118-1124.
- Relyea, R.A., 2012. New Effects of Roundup<sup>®</sup> on Amphibians: Predators Reduce Herbicide Mortality; Herbicides Induce Antipredator Morphology. Ecological Applications 22(2):634-647.
- Richard, S., S. Moslemi, H. Sipahutar, N. Benachour, and G. Seralini, 2005. Differential Effects of Glyphosate and Roundup<sup>®</sup> on Human Placental Cells and Aromatase. Environmental Health Perspectives 113(6):716-720.
- Samsel, A. and S. Seneff, 2013. Glyphosate's Suppression of Cytochrome P450 Enzymes and Amino Acid Biosynthesis by the Gut Microbiome: Pathways to Modern Diseases. Entropy 15:1416-1463.
- Sanchis, J., L. Kantiani, M. Llorca, F. Rubio, A. Ginebreda, J. Fraile, T. Garrido, and M. Farre, 2011. Determination of Glyphosate in Groundwater Samples Using an Ultrasensitive Immunoassay and Confirmation by On-Line Solid-Phase Extraction Followed by Liquid Chromatography Coupled to Tandem Mass Spectrometry. Analytical and Bioanalytical Chemistry 402(7):2335-2345.
- Schummer, C., E. Mothiron, B.M.R. Appenzeller, A. Rizet, R. Wennig, and M. Millet, 2010. Temporal Variations of Concentrations of Currently Used Pesticides in the Atmosphere of Strasbourg, France. Environmental Pollution 158:576-584.
- Scribner, E.A., W.A. Battaglin, J.E. Dietze, and E.M. Thurman, 2003. Reconnaissance Data for Glyphosate, Other Selected Herbicides, Their Degradation Products, and Antibiotics in 51 Streams in Nine Midwestern States, 2002. U.S. Geological Survey Open-File Report 03-217, 101 pp.
- Scribner, E.A., W.A. Battaglin, R.J. Gilliom, and M.T. Meyer, 2007. Concentrations of Glyphosate, Its Degradation Product, Aminomethylphosphonic Acid, and Glufosinate in Ground- and Surface-Water, Rainfall, and Soil Samples Collected in the United States, 2001-2006. U.S. Geological Survey Scientific Investigations Report 2007-5122, 111 pp.
- Skark, C., N. Zullei-Seibert, U. Schottler, and C. Schlett, 1998. The Occurrence of Glyphosate in Surface Water. International Journal of Environmental Analytical Chemistry 70(1-4):93-104.
- Stone, W.W. and J.T. Wilson, 2006. Preferential Flow Estimates to an Agricultural Tile Drain with Implications for Glyphosate Transport. Journal of Environmental Quality 35:1825-1835.
- Struger, J., D. Thompson, B. Staznik, P. Martin, T. McDaniel, and C. Marvin, 2008. Occurrence of Glyphosate in Surface Waters of Southern Ontario. Bulletin of Environmental Contamination and Toxicology 80:378-384.
- U.S. Department of Agriculture, 2009. National Agricultural Statistics Service Charts and Maps for Field Crops. http://www.nass. gov/Charts\_and\_Maps/Field\_Crops/index.asp, accessed October 2009.
- U.S. Department of Agriculture, 2011. Pesticide Data Program Annual Summary, Calendar Year 2009. U.S. Department of Agriculture Agricultural Marketing Service, Science and Technology Programs. http://www.ams.usgs.gov/pdp, *accessed* October 2011.

- U.S. Environmental Protection Agency, 1993. Reregistration Eligibility Decision (RED) Glyphosate. U.S. Environmental Protection Agency Prevention, Pesticides, and Toxics Substances, EPA-738-R-93-014, 290 pp.
- U.S. Environmental Protection Agency, 2002. Occurrence Summary and Use Support Document for the Six-Year Review of National Primary Drinking Water Regulations: Glyphosate. U.S. Environmental Protection Agency, Office of Water, EPA-815-D-02-006, pp. 190-197.
- U.S. Environmental Protection Agency, 2009. Final List of Initial Pesticide Active Ingredients and Pesticide Inert Ingredients to be Screened under the Federal Food, Drug, and Cosmetic Act. Federal Register 74(71):17579-17585.
- U.S. Environmental Protection Agency, 2011. Glyphosate Pesticide Tolerance. Federal Register 76(91):27268-27271.
- Vera, M.S., L. Lagomarsino, M. Sylvester, G.L. Perez, P. Rodriguez, H. Mugni, R. Sinistro, M. Ferraro, C. Bonetto, H. Zagarese, and H. Pizarro, 2010. New Evidence of Roundup<sup>®</sup> (glyphosate formulation) Impact on the Periphyton Community and the Water Quality of Freshwater Ecosystems. Ecotoxicology 19:710-721.
- Vereecken, H., 2005. Mobility and Leaching of Glyphosate: A Review. Pest Management Science 61:1139-1151.
- Vogel, J.R., M.S. Majewski, and P.D. Capel, 2008. Pesticides in Rain in Four Agricultural Watersheds in the United States. Journal of Environmental Quality 37:1101-1115.
- Wan, M.T., J.E. Rahe, and R.G. Watts, 1998. A New Technique for Determining the Sublethal Toxicity of Pesticides to the Vesicular-Arbuscular Mycorrhizal Fungus *Glomus intraradices*. Environmental Toxicology and Chemistry 17(7):1421-1428.
- Williams, B.K. and R.D. Semlitsch, 2010. Larval Responses of Three Midwestern Anurans to Chronic, Low-Dose Exposures of Four Herbicides. Archives of Environmental Contamination and Toxicology 58:819-827.
- Wittmer, I.K., R. Scheidegger, H. Bader, H. Singer, and C. Stamm, 2011. Loss Rates of Urban Biocides Can Exceed Those of Agricultural Pesticides. Science of the Total Environment 409:920-932.
- Young, B.G., 2006. Changes in Herbicide Use Patterns and Production Practices Resulting from Glyphosate-Resistant Crops. Weed Technology 20:301-307.
- Zobiole, L.H.S., R.J. Kremer, R.S. Oliveira, and J. Constantin, 2010. Glyphosate Affects Micro-Organisms in Rhizospheres of Glyphosate-Resistant Soybeans. Journal of Applied Microbiology 110:118-127.